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Using model-data fusion to interpret past trends, and quantify uncertainties in future projections, of terrestrial ecosystem carbon cycling

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Abstract

Uncertainties in model projections of carbon cycling in terrestrial ecosystems stem from inaccurate parameterization of incorporated processes (endogenous uncertainties) and processes or drivers that are not accounted for by the model (exogenous uncertainties). Here, we assess endogenous and exogenous uncertainties using a model-data fusion framework benchmarked with an artificial neural network (ANN). We used 18 years of eddy-covariance carbon flux data from the Harvard forest, where ecosystem carbon uptake has doubled over the measurement period, along with 15 ancillary ecological data sets relative to the carbon cycle. We test the ability of combinations of diverse data to constrain projections of a process-based carbon cycle model, both against the measured decadal trend and under future long-term climate change. The use of high-frequency eddy-covariance data alone is shown to be insufficient to constrain model projections at the annual or longer time step. Future projections of carbon cycling under climate change in particular are shown to be highly dependent on the data used to constrain the model. Endogenous uncertainties in long-term model projections of future carbon stocks and fluxes were greatly reduced by the use of aggregated flux budgets in conjunction with ancillary data sets. The data-informed model, however, poorly reproduced interannual variability in net ecosystem carbon exchange and biomass increments and did not reproduce the long-term trend. Furthermore, we use the model-data fusion framework, and the ANN, to show that the long-term doubling of the rate of carbon uptake at Harvard forest cannot be explained by meteorological drivers, and is driven by changes during the growing season. By integrating all available data with the model-data fusion framework, we show that the observed trend can only be reproduced with temporal changes in model parameters. Together, the results show that exogenous uncertainty dominates uncertainty in future projections from a data-informed process-based model.

Keywords: climate change, data-informed model, Harvard forest, long-term trend, model benchmark, model-data fusion, Monte Carlo, multiple constraints

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Introduction

Terrestrial ecosystems mediate a large portion of the CO_2 flux between the Earth's surface and the atmosphere, with approximately 120 Pg C yr⁻¹ taken up by gross photosynthesis, and a slightly smaller amount respired back (Prentice *et al.*, 2000; Beer *et al.*, 2010; Pan *et al.*, 2011). The balance of these two numbers, net ecosystem exchange (NEE), drives the terrestrial carbon cycle and is tightly coupled to the growth rate of atmospheric CO_2 (Bousquet *et al.*, 2000; Knorr *et al.*, 2007). For policy makers, and many earth-system scientists, a

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major goal of global change research is therefore to understand the processes responsible for changes in terrestrial carbon cycling and to project future states of ecosystems and climate at decadal, or even longer time scales (Clark *et al.*, 2001; Luo *et al.*, 2011).

Increasingly, many long-term data sets show trends that demand investigation. Inventory data show increased forest growth rates in eastern North America (McMahon *et al.*, 2010), potentially due to recent changes in climate, nutrient deposition, or community structure. Similar increases in tropical (Lewis *et al.*, 2009) and temperate (Urbanski *et al.*, 2007; Salzer *et al.*, 2009; Dragoni *et al.*, 2011; Pilegaard *et al.*, 2011) forest carbon uptake have been reported (but see Fahey *et al.*, 2005), and have been linked to changes in the growing season length, and vegetation dynamics. Open questions remain as to the dominant controls of such longterm changes, and the relative importance of climatic and biotic factors (Richardson *et al.*, 2007). As we move into a data-rich era in ecology (Luo *et al.*, 2008), and an era of advanced data mining (e.g., Abramowitz *et al.*, 2007; Moffat *et al.*, 2010) and model uncertainty analysis techniques (e.g., Braswell *et al.*, 2005; Wang *et al.*, 2009; Williams *et al.*, 2009; Keenan *et al.*, 2011c), we are now in a position to address such long-term questions.

Process-based models are the most commonly used tools for the projection of long-term ecosystem function. For terrestrial vegetation, the term 'process-based' incorporates a broad range of methodologies for describing eco-physiological processes, from semi-empirical relationships to mechanistic descriptions based on physical laws. Such models are often shown to reproduce observations 'reasonably well' (e.g., Braswell *et al.*, 2005; Williams *et al.*, 2005). However, model intercomparisons and model-data comparison studies show tremendous variations among models for both short- and long-term projections (e.g., Friedlingstein *et al.*, 2006; Siqueira *et al.*, 2006; Sich *et al.*, 2008; Schwalm *et al.*, 2010; Dietze *et al.*, 2011; Keenan *et al.*, in press).

Model-data fusion (also referred to as 'data assimilation', or 'inverse modeling') (Wang et al., 2009; Keenan et al., 2011c) is a means by which to use observational data to optimize a model and quantify model uncertainty. The approach identifies combinations of model parameters that give an equivalent model-data agreement. In this way, data from different sources can be synthesized using the model as the interpreter, independent of parameter assumptions. Results are conditional on model structure, and the information content of observational data along with data uncertainties (Raupach et al., 2005; Keenan et al., 2011c). For example, model-data fusion applications of both simple (Braswell et al., 2005) and complex (Medvigy et al., 2009) models at Harvard forest acknowledged the limitation of using only one or two data streams to constrain model parameterization.

Even with an optimized model, results remain contingent on model structure. An optimized model is therefore not necessarily correct or even good. For example, if the model structure is inadequate, or the model parameters are not well constrained, an optimized model can get the right answer for the wrong reason or through a variety of unverified process combinations (equifinality) (Beven, 2006). It is thus important to test the optimized model against data that was not used for training. Another approach to assessing model performance is to test the optimized model using an independent 'benchmark'. Empirical data-mining tools such as artificial neural networks (ANN) can serve as an excellent means by which to benchmark model performance (Abramowitz *et al.,* 2007). Such data-mining tools have been shown to capture the complex response of ecosystem carbon cycling to climatic drivers (Moffat *et al.,* 2010). They therefore provide an indication of how well a good (though not necessarily best) model should be expected to perform.

Carbon uptake at Harvard forest has increased from ~200 to ~500 g C m⁻² yr⁻¹ during the 18-year period from 1992 to 2009; around this long-term trend, there is also interannual variability on the order of ±117 g $C m^{-2} yr^{-1}$ (1 SD). In this paper, we use a parsimonious forest carbon cycle model, embedded in a multiple constraints Markov-chain Monte Carlo optimization framework, to examine trends and variability in uptake. We first assess the impact of using different data constraints on uncertainty in model performance, both in training and test periods. An ANN approach (Moffat et al., 2010) is then used to benchmark the optimized process-based model. By examining how the use of different constraints can reduce uncertainty, we test whether recent changes in uptake are driven by concurrent trends external to the model system (exogenous factors) or model-internal (endogenous) factors. The impact of endogenous uncertainty in ecological forecasting is also assessed and compared with current trends in carbon uptake at the Harvard forest.

Materials and methods

Site

All data used were obtained within the footprint of the eddycovariance tower at the Harvard Forest Environmental Measurement Site (HFEMS) (http://atmos.seas.harvard.edu/lab/ hf/index.html), which is located in the New England region of the northeastern United States (42.53 N 72.17 W, elevation 340 m) (Wofsy *et al.*, 1993; Barford *et al.*, 2001; Urbanski *et al.*, 2007). The forest within the tower footprint is largely deciduous, dominated by red oak (*Quercus rubra*, 52% basal area), red maple (*Acer rubrum*, 22% basal area), eastern hemlock (*Tsuga canadensis*, 17% basal area), and a secondary presence of white pine (*Pinus strobus*) and red pine (*Pinus resinosa*) is also found within the tower footprint.

Data

We used 18 complete years (1992–2009) of hourly meteorological and eddy-covariance (Wofsy *et al.*, 1993; Goulden *et al.*, 1996; Barford *et al.*, 2001; Urbanski *et al.*, 2007) measurements of NEE (http://atmos.seas.harvard.edu/lab/data/nigec-data. html). Hourly gap-filled meteorological variables used include incident photosynthetically active radiation (PAR), air temperature above the canopy, soil temperature at a depth of 5 cm, vapor pressure deficit (VPD), and atmospheric CO_2 concentration. Quality controlled hourly eddy-covariance observations (without gap-filling) of NEE were used to optimize the ecosystem model and train the ANN. Gap-filled NEE values were only used to provide annual sums for evaluating optimized model performance.

For ancillary data constraints, we used measurements of leaf area index (LAI), soil organic carbon content, carbon in roots, carbon in wood, wood carbon annual increment, observer-based estimates of bud-burst and leaf senescence, leaf litter, woody litter, and continuous and manual measurements of soil respiration (Table 1), downloaded from the Harvard forest data repository (http://harvardforest.fas.harvard.edu/ data/archive.html).

In addition to the ancillary data available from the Harvard forest data repository, we used two other model constraints: (1) annual estimates of the contribution of root respiration to total soil respiration and (2) estimates of turnover times of soil organic matter pools. Radiocarbon and soda-lime (in combination with trenching) based estimates of the contribution of autotrophic respiration (Ra) to total soil respiration (Rsoil) were obtained from Gaudinski et al. (2000), Bowden et al. (1993), and E. Davidson (unpublished results). Bowden et al. (1993) provided a mean annual estimate of belowground autotrophic respiration as roughly 33% of total annual soil respiration. Gaudinski et al. (2000) and E. Davidson (unpublished results) suggested an approximate error of roughly 50% associated with this estimate. Although annual fluxes were constrained to a specific proportion, Ra: Rsoil could vary on shorter timescales. Turnover times of litter and the two soil organic matter pools (slow, passive) were also taken from Gaudinski et al. (2000). Microbial biomass turnover times were estimated as 1.7 ± 1.3 years (E. Davidson unpublished results).

Estimates of uncertainty were used for each data stream in the optimization. Uncertainty estimates for NEE were taken from Richardson et al. (2006), where uncertainties were shown to follow a double-exponential distribution, with the standard deviation of the distribution specified as a linear function of the flux. Estimates of uncertainty due to flux gap-filling (which apply to the annual NEE totals) were taken from Barr et al. (2009). Soil respiration uncertainty estimates were taken from Savage et al. (2009) and Phillips et al. (2010), where measurement uncertainty increased linearly with the magnitude of the flux. LAI sampling uncertainties were estimated as the standard error (n = 34 plots) of the mean LAI. Litterfall sampling errors were calculated as the standard error (n = 34)plots) of the annual total litterfall across all plots. Uncertainty of carbon in wood was calculated from the standard error (n = 34 plots, 635 trees) of the mean plot-level cumulative increment, which averaged ~10% over all years. Two independent measurements (Bowden et al., 1993; Gaudinski et al., 2000) were used to constrain the initial value of total soil C content (C_{SOM} = 8.3 ± 1.4 kg C m⁻²; mean ± 1 SE), with uncertainties estimated based on the standard deviation between datasets. Root biomass uncertainties were estimated from spatial variation in the samples (n = 21 plots), taken in the control plots of the DIRT project (http://www.lsa.umich. edu/eeb/labs/knute/DIRT/). Uncertainty estimates for the dating of phenological events were based on the between tree standard deviation.

Additionally, three different soil respiration data sets, two automated and one manual, were used (Savage *et al.*, 2009; Phillips *et al.*, 2010). Although seasonal cycles were similar between the data sets, disagreement in the magnitude of the flux was evident between the different soil respiration data sets, reflecting high spatial variability in soil characteristics. We included three additional scaling parameters (data

Measurement	Frequency	No. of data points	Reference
Eddy-covariance	Hourly	73 198	Urbanski et al. (2007) and *
Soil respiration 1	Hourly	26 430	Savage <i>et al.</i> (2009)
Soil respiration 2	Hourly	19 030	Phillips et al. (2010)
Soil respiration 3	Weekly	498	Ť
Leaf area index	Monthly	51	Norman (1993), Urbanski $et~al.$ (2007), and *
Leaf litter fall	Yearly	10	Urbanski <i>et al.</i> (2007) and [*]
Woody biomass	Yearly	15	Jenkins et al. (2004), Urbanski et al. (2007), and *
Woody litterfall	Yearly	8	Urbanski <i>et al.</i> (2007) and [*]
Root biomass	1 year	1	DIRT project [*]
Forest floor carbon	1 year	1	Gaudinski et al. (2000)
Budburst	Yearly	15	O'Keefe (2000) [*]
Leaf drop	Yearly	14	O'Keefe (2000)*
Soil carbon pools	3 years	3	Gaudinski <i>et al.</i> (2000), Magill <i>et al.</i> (2000), Bowden <i>et al.</i> (1993)
Soil carbon turnover	One	1	Gaudinski et al. (2000)
Proportion of heterotrophic respiration in soil	One	1	Gaudinski et al. (2000)

Table 1 Data sets used in this study	Table 1	used in this study
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*See data download page: http://harvardforest.fas.harvard.edu/data/archive.html. †ftp://ftp.as.harvard.edu/pub/nigec/HU_Wofsy/hf_data/ecological_data/soilR/. harmonizing parameters) in the optimization process (e.g., van Oijen *et al.*, 2011). These scale different chamber datasets to account for the possibility that a particular dataset is not representative of the mean soil respiration of the tower footprint. This thus harmonizes the magnitude of the different soil respiration data streams to give an estimate of the spatial average soil respiration of the tower footprint, but then leverages the temporal patterns in the data as model constraints.

The FöBAAR model

We developed a forest carbon cycle model that strikes a balance between parsimony and detailed process representation. Working on an hourly timescale, FöBAAR (Forest Biomass, <u>Assimilation, Allocation and Respiration</u>) calculates photosynthesis from two canopy layers, and respiration from eight carbon pools [leaf, wood, roots, soil organic matter (microbial, slow and passive pools), leaf litter and (during phenological events) mobile stored carbon], using as environmental forcings canopy air temperature (T_a), 5 cm soil temperature (T_s), photosynthetic active radiation (PAR), VPD, and atmospheric CO₂.

The canopy in FöBAAR is described in two compartments representing sunlit and shaded leaves (Sinclair et al., 1976; Wang & Leuning, 1998). Intercepted radiation by sunlit or shade leaves depends on the position of the sun, and the area of leaf exposed to the sun based on leaf angle and the canopy's ellipsoidal leaf distribution (Campbell, 1986). Here, we assume a spherical leaf angle distribution. Assimilation rates for sunlit and shaded leaves are calculated through the commonly used Farquhar approach (Farquhar et al., 1980; De Pury & Farquhar, 1997), with dependencies on absorbed direct and diffuse radiation, air temperature, VPD, and the concentration of CO₂ within the leaf inter-cellular spaces. Stomatal conductance is calculated using the Ball-Berry model (Ball et al., 1987), coupled to photosynthetic rates through the analytical solution of the Farquhar, Ball Berry coupling (Baldocchi, 1994). Rates of photosynthesis are dependent on the minimum between rate of carboxylation and the proportional rate of electron transport. The canopy integrated (over space and time) RuBP (ribulose-1,5-bisphosphate) rate of carboxylation, V_c , and the rate of electron transport, I, are calculated following Farquhar et al. (1980) and De Pury & Farquhar (1997). The CO₂ compensation point and the mitochondrial respiration rate are calculated using an Arrhenius-type equation (Bernacchi et al., 2001).

Maintenance respiration is calculated as a fraction of assimilated carbon. The remaining assimilate is allocated to foliar carbon, then to the wood and root carbon pools on a daily time step. Mobile stored carbon relates only to foliage and is respired only during periods of bud-burst and leaf-fall. Carbon allocation and canopy phenology are simulated as in the DALEC model (Williams *et al.*, 2005; Fox *et al.*, 2009).

Root respiration is calculated hourly and coupled to photosynthesis through the direct allocation to roots. Dynamics of soil organic matter is modeled using a three-pool approach (microbial, slow, and passive pools) (Knorr & Kattge, 2005). Decomposition in each pool is calculated hourly, with a pool specific temperature dependency. Litter decomposition is also calculated hourly, but on an air temperature basis. Litter and root carbon are transferred to the microbial pool, then to the slow and finally to the passive pool.

In total, 35 model parameters (including three data harmonization parameters, Table 2; P40, P41, P42) and seven initial pools were optimized, giving a total of 42 free parameters. The inclusion of the initial biomass and soil pools in the optimization process removed the need for a model spin-up.

Model-data fusion

An adaptive multiple constraints Markov-chain Monte Carlo (MC₃) optimization was used to optimize the process-based model and explore model uncertainty. The algorithm uses the Metropolis–Hastings (M-H) approach (Metropolis & Ulam, 1949; Metropolis *et al.*, 1953; Hastings, 1970) combined with simulated annealing (Press *et al.*, 2007). It is loosely based on that of Braswell *et al.* (2005), and it is adaptive in the sense that the step size, which is expressed as a fraction of the initial parameter range, is automatically adjusted to obtain a fixed acceptance rate. Preliminary tests with synthetic data indicated an acceptance rate of ~21% gave optimal efficiency (good mixing) for the posterior exploration. Prior distributions for each parameter given in Table 2 were assumed to be uniform (noninformative, in a Bayesian context).

The optimization process uses a two-step approach. In the first stage, the parameter space is explored for 100 000 iterations using the MC_3 optimization algorithm. At each iteration, the current step size is used as the standard deviation of random draws from a normal distribution with mean zero, by which parameters are varied around the previous accepted parameter set. Parameters that fall outside the initial parameter range are 'bounced' back within their range. This stage identifies the optimum parameter set by minimizing the cost function [see Eqn (2)], and 100 000 model iterations were used to identify the optimum parameter set, as longer runs led to no improvement.

In the second stage, the parameter space is again explored, and a parameter set is accepted if the cost function for each data stream (defined below) passes a χ^2 test (at 90% confidence) for acceptance/rejection (after variance normalization based on the minimum cost function obtained (e.g., Franks *et al.*, 1999; Richardson *et al.*, 2010). This approach is preferable to using the aggregate cost function, as it ensures that model predictions are consistent with each of the individual data streams.

The cost function quantifies the extent of model-data mismatch using all available data (eddy-covariance, biometric, etc.), constructed here as in Keenan *et al.* (2011c). Individual data stream cost functions, j_i , are calculated as the total uncertainty-weighted squared data-model mismatch, averaged by the number of observations for each data stream (N_i):

$$j_{i} = \left(\sum_{t=1}^{N_{i}} \left(\frac{y_{i}(t) - p_{i}(t)}{\delta_{i}(t)}\right)^{2}\right) / N_{i},$$
(1)

where $y_i(t)$ is a data constraint at time t for data stream i and $p_i(t)$ is the corresponding model predicted value. $\delta_i(t)$ is the measurement specific uncertainty. For the aggregate multiobjective cost function, we use the average of the individual cost functions, which can be written as follows: **Table 2** FöBAAR model parameters and pools. Both parameters and initial pool sizes were optimized conditional on the dataconstraints. The posterior 90% confidence interval for each parameter is given, based on optimization to Period 2 using all dataconstraints

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Id	Name	Definition	Min	Max	90% CI		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$				Initial cark	ial carbon pools (g C m ^{-2})			
P2 V_{cc} Carbon in large 8 000 1 4 000 7 792, 19 91 P3 Like, Carbon in slow cycling soil organic matter layer 10 1 000 95, 278 P4 SOM _c passive Carbon in in passive cycling soil organic matter layer 10 1 000 95, 278 P5 SOM _c passive Carbon in passive cycling soil organic matter layer 1500 12 000 1800, 4560 P6 Mobie After Fraction of GPP allocated to foliage 0.1 1 0.31, 0.48 P7 Af Fraction of GPP allocated to roots 0.5 1 0.57, 0.83 P10 Lif, V Litterfall from wood (Log ₁₀) -6 -1 -2.42, -1.88 P11 Lifs Fraction of Cf of transferred to mobile carbon 0.3 0.8 0.03, 0.07 P13 LifsSOM Ta Litter to slow SOM _c transferr ate (Log ₁₀) -6 -1 -2.79, -2.19 P14 LidgSOM Ta Slow SOM _c to passive SOM _c rate 0.3 0.8 0.03, 0.55 SOM ₆₂ SOM _p Ta Slow SOM _c to passive SOM _c rate 0.01	P1	R _C	Carbon in roots	20	500	28, 205		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P2	W _C	Carbon in wood	8 000	14 000	7 792, 10 931		
P4 SOM _C slow Carbon in slow cycling soil organic matter layer 10 100 95, 278 P5 SOM _C passive Carbon in spassive cycling soil organic matter layer 1500 12 000 90, 175 P7 Af Fraction of CPP allocated to foliage 0.1 1 0.31, 0.48 P8 Ar Fraction of CPP allocated to roots 0.5 1 0.57, 0.83 P9 Lf ₄ Litterfall from roots (O_{00}_{0}) -6 -1 -1.12, -0.88 P11 Lf ₄ Litterfall from roots (O_{00}_{0}) -6 -1 -2.62, -1.88 P13 Lit ₅ OM T ₄ Litter to slow SOM _C to passive SOM _C rate 0.03 0.8 0.03, 0.05 P14 Lit ₅ SOM ₇ Slow SOM _C to passive SOM _C rate 0.01 0.07, 0.07 0.15 P15 SOM ₆₅ SOM ₇ Slow SOM _C to passive SOM _C rate 0.03 0.8 0.03, 0.55 Canopy parameters P17 LMA Leaf mass per area (g C m ⁻⁵) 50 120 81, 120 180 800, 250 000 80, 007, 5000 820, 007, 50	Р3	Litc	Carbon in litter	10	1 000	146, 528		
P5 SOM_passive Mobic Carbon in passive cycling soil organic matter layer Mobile carbon 1500 12 000 1800, 4 560 P6 Mobic Mobile carbon 75 200 90, 175 P7 Af Fraction of NP allocated to roots 0.1 1 0.31, 0.48 P8 Ar Fraction of NP allocated to roots 0.5 1 0.57, 0.83 P10 Lif Litterfall from (oliging (0.5g ₁₀) -6 -1 -2.12, -0.88 P11 Lif Litterfall from toots (0.5g ₁₀) -6 -1 -2.69, -1.12, -0.88 P12 Fc_If Fraction of Cf not transferred to mobile carbon 0.3 0.8 0.36, 0.52 P13 LitgSOM T _d Slow SOM _c to passive SOM _c rate 0.01 0.8 0.07, 0.77 P15 SOM _a SOM _p T _d Slow SOM _c to passive SOM _c rate 0.01 0.8 0.03, 0.8 0.07, 0.07 P14 LitgSOM T _d Leaf mass per arca (g C m ⁻²) 50 100 1.80, 0.55 Canopy parameters P14 LitgSOM T _d Leaf mass per	P4	SOM_C slow	Carbon in slow cycling soil organic matter layer	10	1 000	95,278		
P6 Mobile carbon 75 200 90, 175 P7 Af Fraction of GPP allocated to foliage 0.1 1 0.31, 0.48 P8 Ar Fraction of NPP allocated to roots 0.5 1 0.57, 0.83 P9 Lf _w Litterfall from foliage (Log ₁₀) -6 -1 -5.14, -4.48 P11 Lf _w Litterfall from roots (Log ₁₀) -6 -1 -2.62, -1.88 P12 Fc_lf Fraction of Cf not transferred to mobile carbon 0.3 0.8 0.36, 0.52 P13 Lit_SOM T _a Litter to slow SOM _c transferrate (Log ₁₀) -6 -1 -2.79, -2.09 P14 Lit_SOM T _a Slow SOM _C to passive SOM _c rate 0.01 0.8 0.03, 0.8 0.36, 0.52 P15 SOM ₆₂ SOM _P T _a Slow SOM _c to passive SOM _c rate 0.01 0.8 0.07, 0.77 P16 SOM ₆₂ SOM _P T _a Slow SOM _c to passive SOM _c rate 0.01 0.8 0.03, 0.80 0.050 0.00 2.00 0.20 0.00, 2.00 0.00 2.00 0.00 50 0.01 0.5 0.01 0.5 0.00, 2.50<	P5	SOM _C passive	Carbon in passive cycling soil organic matter laver	1 500	12 000	1 800, 4 560		
Allocation and Transfer parameters P7 Af Fraction of GPP allocated to foliage 0.1 1 0.31, 0.48 P8 Ar Fraction of NPP allocated to roots 0.5 1 0.57, 0.83 P1 Lift Litterfall from toolinge (Log ₁₀) -6 -1.12, -0.88 P10 Liw Litterfall from roots (Log ₁₀) -6 -1 -2.42, -1.88 P11 Lift Litter foll from roots (Log ₁₀) -6 -1 -2.79, -2.09 P13 LitsSOM 7 _d Litter to slow SOM _c transfer rate (Log ₁₀) -6 -1 -2.79, -2.09 P14 LitsSOM 7 _d Slow SOM _c to passive SOM _c rate 0.01 0.5 0.01, 0.07 P15 SOM _{s2} SOM _p 7 _d Slow SOM _c to passive SOM _c temp. dependence 0.01 0.8 0.03, 0.55 Canopy parameters P17 LMA Leaf mass per area (g C m ⁻²) 150 60 180, 550 P19 Vemax Activation energy for Vemax 200 000 200 000 200 000, 200 00 200 000, 200 00 200 000, 200 00	P6	Mobe	Mobile carbon	75	200	90, 175		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$				Allocation and transfer parameters				
P8 Ar Fraction of NPP allocated to roots 0.5 1 0.57, 0.83 P10 Lft Litterfall from foliage (\log_{10}) -6 -0.85 -1.12, -0.88 P10 Lft Litterfall from roots (\log_{10}) -6 -1 -2.62, -1.88 P11 Lft Litterfall from roots (\log_{10}) -6 -1 -2.62, -1.88 P13 LittySOM Litter to slow SOM_c transfer rate (\log_{10}) -6 -1 -2.79, -2.09 P14 LittySOM Td Litter to slow SOM_c transfer rate (\log_{10}) -6 -1 -2.79, -2.09 P15 SOMs_SOM_T Z Slow SOM_c to passive SOM_c rate 0.03 0.8 0.07, 077 P16 SOMs_SOM_T Z Slow SOM_c to passive SOM_c rate 0.01 0.8 0.03, 0.55 P17 LMA Leaf mass per area (g C m^{-2}) 50 60 180, 050 175 09, 165 P18 MaxFol Maximum canopy carbon content (g C m^{-2}) 50 60 180, 050 200, 000 200, 000, 250, 000 200, 000, 250, 000 200, 000, 250, 000 200, 000, 250, 000 200, 000, 250, 000 200, 000, 250, 000 200, 000	P7	Af	Fraction of GPP allocated to foliage	0.1	1	0.31, 0.48		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P8	Ar	Fraction of NPP allocated to roots	0.5	1	0.57, 0.83		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P9	Lf_{f}	Litterfall from foliage (Log ₁₀)	-6	-0.85	-1.12, -0.88		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P10	Lf_w	Litterfall from wood (Log ₁₀)	-6	-1	-5.14, -4.88		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P11	Lf _r	Litterfall from roots (Log_{10})	-6	-1	-2.62, -1.88		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P12	Fc_lf	Fraction of Cf not transferred to mobile carbon	0.3	0.8	0.36, 0.52		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	P13	Lit ₂ SOM	Litter to slow SOM_C transfer rate (Log ₁₀)	-6	-1	-2.79, -2.09		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P14	$Lit_2SOM T_d$	Litter to slow SOM _C temperature dependence	0.01	0.5	0.01, 0.07		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P15	SOM _{S2} SOM _P	Slow SOM _C to passive SOM _C rate	0.03	0.8	0.07, 0.77		
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	P16	$SOM_{s2}SOM_{P}T_{d}$	Slow SOM _C to passive SOM _C temp. dependence	0.01	0.8	0.03, 0.55		
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$				Canopy parameters				
P18 MaxFol Maximum canopy carbon content (g C m ⁻²) 150 600 180, 550 P19 Vc _{max} Velocity of carboxylation (umol mol ⁻¹) 60 175 90, 165 P20 Ea Vc _{max} Deactivation energy for Vc _{max} 58 000 75 000 280 000, 250 000 P21 Ed Vc _{max} Deactivation energy for Vc _{max} 200 000 250 000 40 000, 50 000 P23 Ed J _{max} Activation energy for the electron transport rate 40 000 50 000 180 000, 230 000 P24 Rd Rate of dark respiration 0.01 1.1 0.01, 1.1 P25 Q10 Rd Temperature dependence of Rd 0.4 2.8 0.45, 2.75 P17 GDD ₁ Growing degree day initiation 50 150 91, 117 P27 GDD ₁ Growing degree days for spring onset 135 300 135, 277 P28 Air T _s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P39 Lit _d Litter respiration rate (Log ₁₀) -7 -1 $-6.6, -3.7$ P31 Lit _d Litter resp	P17	LMA	Leaf mass per area (g C m ⁻²)	50	120	81, 120		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	P18	MaxFol	Maximum canopy carbon content (g C m ^{-2})	150	600	180, 550		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	P19	Vc _{max}	Velocity of carboxylation (umol mol^{-1})	60	175	90, 165		
P21 Ed Vc_max Deactivation energy for Vc_max 200 000 250 000 200 000, 250 000 P22 Ed J_max Activation energy for the electron transport rate 40 000 50 000 40 000, 50 000 P23 Ed J_max Deactivation energy for the electron transport rate 40 000 50 000 40 000, 50 000 P23 Ed J_max Deactivation energy for the electron transport rate 180 000 230 000 180 000, 230 000 P24 Rd Rate of dark respiration 0.01 1.1 0.01, 1.1 P25 Q10 Rd Temperature dependence of Rd 0.4 2.8 0.45, 2.75 Phenology parameters Sol 150 91, 117 P27 GDD1 Growing degree days for spring onset 135 300 135, 277 P28 Air T_s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P29 GDD2 Spring photosynthetic GDD maximum 70 -7 -1 -66, -3.7 P31 Litd, T_d Litter respiration rate (Log_{10}) -6 -1 -455, 3.11 P33 SOMsd Slow cycling	P20	Ea Vc _{max}	Activation energy for Vc _{max}	58 000	75 000	58 000,75 000		
P22 Ea J_{max} Activation energy for the electron transport rate 40 000 50 000 40 000, 50 000 P23 Ed J_{max} Deactivation energy for the electron transport rate 180 000 230 000 180 000, 230 000 P24 Rd Rate of dark respiration 0.01 1.1 0.01, 1.1 P25 Q10 Rd Temperature dependence of Rd 0.4 2.8 0.45, 2.75 Phenology parameters 50 150 91, 117 P27 GDD ₁ Growing degree days for spring onset 135 300 135, 277 P28 Air T _s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P29 GDD ₂ Spring photosynthetic GDD maximum -77 -1 $-6.6, -3.7$ P31 Lit _d Litter respiration rate (Log ₁₀) -77 -1 $-6.3, -5.3, 3.11$ P33 SOM _{5d} T _d Slow cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-6.38, -5.15$ P34 SUM _{5d} T _d Slow cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-6.38, -5.15$ P33 SOM _{5d} T _d	P21	Ed Vc _{max}	Deactivation energy for Vc _{max}	200 000	250 000	200 000, 250 000		
P23 Ed J_{max} Deactivation energy for the electron transport rate 180 000 230 000 180 000, 230 000 P24 Rd Rate of dark respiration 0.01 1.1 0.01, 1.1 P25 Q10 Rd Temperature dependence of Rd 0.4 2.8 0.45, 2.75 P26 GDD ₀ Day of year for growing degree day initiation 50 150 91, 117 P27 GDD ₁ Growing degree days for spring onset 135 300 135, 277 P28 Air T _s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P29 GDD ₂ Spring photosynthetic GDD maximum 500 1 000 660, 1 000 Respiration parameters 77 -1 $-6.6, -3.7$ -6 -1 $-4.55, 3.11$ P30 Lit _d Litter respiration rate (Log ₁₀) -6 -1 $-4.55, 3.11$ P33 SOM _{Sd} Slow cycling SOM _C temperature dependence 0.01 0.2 $0.01, 0.1$ P34 SOM _{Pd} Passive cycling SOM _C temperature dependence 0.01 0.2 $0.07, 0.2$ P35 <td>P22</td> <td>Ea J_{max}</td> <td>Activation energy for the electron transport rate</td> <td>40 000</td> <td>50 000</td> <td>40 000, 50 000</td>	P22	Ea J _{max}	Activation energy for the electron transport rate	40 000	50 000	40 000, 50 000		
P24 Rd Rate of dark respiration 0.01 1.1 0.01, 1.1 P25 Q10 Rd Temperature dependence of Rd 0.4 2.8 0.45, 2.75 P26 GDD ₀ Day of year for growing degree day initiation 50 150 91, 117 P27 GDD ₁ Growing degree days for spring onset 135 300 135, 277 P28 Air T _s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P29 GDD ₂ Spring photosynthetic GDD maximum -7 -1 $-6.6, -3.7$ P31 Lit _d Litter respiration rate (Log ₁₀) -7 -1 $-6.6, -3.7$ P33 SOM _{sd} Slow cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-4.55, 3.11$ P34 SOM _{Pd} Passive cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-6.38, -5.15$ P35 Rroot _d Root respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P34 SOM _{Pd} Passive cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P35 Rroot _d <td>P23</td> <td>Ed J_{max}</td> <td>Deactivation energy for the electron transport rate</td> <td>180 000</td> <td>230 000</td> <td>180 000, 230 000</td>	P23	Ed J _{max}	Deactivation energy for the electron transport rate	180 000	230 000	180 000, 230 000		
P25 Q10 Rd Temperature dependence of Rd 0.4 2.8 $0.45, 2.75$ P26 GDD ₀ Day of year for growing degree day initiation 50 150 $91, 117$ P27 GDD ₁ Growing degree days for spring onset 135 300 $135, 277$ P28 Air T_s Leaf senescence onset mean air temperature (°C) 0 15 $11, 12.4$ P29 GDD ₂ Spring photosynthetic GDD maximum 500 1000 $660, 1000$ Respiration parameters 200 1000 $660, 1000$ Respiration parameters P30 Lit _d Litter respiration rate (Log ₁₀) -7 -1 $-6.6, -3.7$ P31 Lit _d T _d Litter respiration rate (Log ₁₀) -6 -1 $-4.55, 3.11$ P33 SOM _{sd} T _d Slow cycling SOM _c respiration rate (Log ₁₀) -6 -1 $-6.38, -5.15$ P35 Rroot _d Root respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P34 SOM _{Fd} Passive cycling SOM _c respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P36 <td>P24</td> <td>Rd</td> <td>Rate of dark respiration</td> <td>0.01</td> <td>1.1</td> <td>0.01, 1.1</td>	P24	Rd	Rate of dark respiration	0.01	1.1	0.01, 1.1		
P26GDD0Day of year for growing degree day initiation $\overline{50}$ 150 $91, 117$ P27GDD1Growing degree days for spring onset 135 300 $135, 277$ P28Air T_s Leaf senescence onset mean air temperature (°C) 0 15 $11, 12.4$ P29GDD2Spring photosynthetic GDD maximum 500 1000 $660, 1000$ Respiration parameters 77 -1 $-6.6, -3.7$ P30Lit _d Litter respiration rate (Log ₁₀) -77 -1 $-6.6, -3.7$ P31Lit _d T_d Litter respiration temperature dependence 0.001 0.1 $0.01, 0.1$ P32SOM _{Sd} Slow cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-4.55, 3.11$ P33SOM _{Sd} T_d Slow cycling SOM _C respiration rate (Log ₁₀) -6 -1 $-6.38, -5.15$ P35Rroot _d Root respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P36Rroot _d T_d Root respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P37Mob _C r,Mobile stored carbon respiration rate (Log ₁₀) -6 -1 $-5.09, -3.77$ P38Mob _C T,Fraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ Scaling parametersSoil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P40Rsoil ₁ Soil respiration scaling co-efficient (data set 2) 0.5 2 $0.96, 1.65$ P42Rsoil ₃ Soil res	P25	Q10 Rd	Temperature dependence of Rd	0.4	2.8	0.45, 2.75		
P26 GDD ₀ Day of year for growing degree day initiation 50 150 91, 117 P27 GDD ₁ Growing degree days for spring onset 135 300 135, 277 P28 Air T_s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P29 GDD ₂ Spring photosynthetic GDD maximum 500 1 000 660, 1 000 Respiration Parameters Parameters Parameters -7 -1 -6.6, -3.7 P31 Lit _d T _d Litter respiration temperature dependence 0.001 0.1 001, 0.1 P32 SOM _{Sd} Slow cycling SOM _C respiration rate (Log ₁₀) -6 -1 -4.55, 3.11 P33 SOM _{Sd} T _d Slow cycling SOM _C temperature dependence 0.01 0.2 0.01, 0.19 P34 SOM _{Fd} Passive cycling SOM _C respiration rate (Log ₁₀) -6 -1 -6.38, -5.15 P35 Rroot _d Root respiration rate (Log ₁₀) -6 -1 -5.09, -3.77 P36 Rroot _d T _d Root respiration rate (Log ₁₀) -6 -1.5, 0.5 0.5 P38				Phenology parameters				
P27 GDD1 Growing degree days for spring onset 135 300 135, 277 P28 Air T_s Leaf senescence onset mean air temperature (°C) 0 15 11, 12.4 P29 GDD2 Spring photosynthetic GDD maximum 500 1 000 660, 1 000 Respiration parameters P30 Litd Litter respiration rate (Log10) -7 -1 $-6.6, -3.7$ P31 Litd Td Litter respiration temperature dependence 0.001 0.1 0.01, 0.1 P32 SOMsd Slow cycling SOM _C respiration rate (Log10) -6 -1 $-4.55, 3.11$ P33 SOMsdTd Slow cycling SOM _C temperature dependence 0.01 0.2 0.01, 0.19 P34 SOM _{Pd} Passive cycling SOM _C respiration rate (Log10) -6 -1 $-5.39, -3.77$ P35 Rroot _d Root respiration rate (Log10) -6 -1 $-5.09, -3.77$ P36 Rroot _d Root respiration rate (Log10) -6 -1 $-5.09, -3.77$ P38 Mob _C r Mobile stored carbon respiration rate (Log10) -6 -0.5 $-1.5, 0.5$ <td>P26</td> <td>GDD₀</td> <td>Day of year for growing degree day initiation</td> <td>50</td> <td>150</td> <td>91, 117</td>	P26	GDD ₀	Day of year for growing degree day initiation	50	150	91, 117		
P28Air T_s Leaf senescence onset mean air temperature (°C)01511, 12.4P29GDD2Spring photosynthetic GDD maximum5001 000660, 1 000Respiration parametersP30LitdLitter respiration rate (Log10) -7 -1 $-6.6, -3.7$ P31LitdTdLitter respiration temperature dependence0.0010.10.01, 0.1P32SOMsdSlow cycling SOMc respiration rate (Log10) -6 -1 $-4.55, 3.11$ P33SOMsdTdSlow cycling SOMc temperature dependence0.010.20.01, 0.19P34SOMFdPassive cycling SOMc respiration rate (Log10) -6 -1 $-6.38, -5.15$ P35RrootdRoot respiration rate (Log10) -6 -1 $-5.09, -3.77$ P36RrootdTdRoot respiration rate temperature dependence 0.01 0.2 $0.07, 0.2$ P37MobcrMobile stored carbon respiration rate (Log10) -6 $-1.5, 0.5$ $-1.5, 0.5$ P38MobcrFraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ P39MaintrSoil respiration scaling co-efficient (data set 1) 0.5 2 $0.62, 1.53$ P40Rsoil1Soil respiration scaling co-efficient (data set 2) 0.5 2 $0.45, 1.65$ P41Rsoil3Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P27	GDD_1	Growing degree days for spring onset	135	300	135, 277		
P29GDD2Spring photosynthetic GDD maximum5001 000660, 1 000P30LitdLitter respiration rate (Log10) -7 -1 $-6.6, -3.7$ P31LitdTdLitter respiration temperature dependence 0.001 0.1 $0.01, 0.1$ P32SOMsdSlow cycling SOMc respiration rate (Log10) -6 -1 $-4.55, 3.11$ P33SOMsdTdSlow cycling SOMc temperature dependence 0.01 0.2 $0.01, 0.1, 0.1$ P34SOMpdPassive cycling SOMc respiration rate (Log10) -6 -1 $-6.38, -5.15$ P35RrootdRoot respiration rate (Log10) -6 -1 $-5.08, -5.15$ P36RrootdTdRoot respiration rate temperature dependence 0.01 0.2 $0.07, 0.2$ P37MobcrMobile stored carbon respiration rate (Log10) -6 -0.5 $-1.5, 0.5$ P38MobcrTrFraction of mobile transfers respired 0 0.1 $0, 0.1$ P39MaintrFraction of GPP respired for maintenance 0.5 2 $0.96, 1.65$ P40Rsoil_1Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P42Rsoil_3Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P28	Air $T_{\rm s}$	Leaf senescence onset mean air temperature (°C)	0	15	11, 12.4		
P30Lit $_{d}$ Litter respiration rate (Log $_{10}$)Respiration parametersP31Lit $_{d}T_{d}$ Litter respiration rate (Log $_{10}$) -7 -1 $-6.6, -3.7$ P31Lit $_{d}T_{d}$ Litter respiration temperature dependence 0.001 0.1 $0.01, 0.1$ P32SOM _{5d} Slow cycling SOM _C respiration rate (Log $_{10}$) -6 -1 $-4.55, 3.11$ P33SOM _{5d} T _d Slow cycling SOM _C temperature dependence 0.01 0.2 $0.01, 0.19$ P34SOM _{Pd} Passive cycling SOM _C respiration rate (Log $_{10}$) -6 -1 $-6.38, -5.15$ P35Rroot $_{d}$ Root respiration rate (Log $_{10}$) -6 -1 $-5.09, -3.77$ P36Rroot $_{d}T_{d}$ Root respiration rate (Log $_{10}$) -6 -1 $-5.09, -3.77$ P37Mob _C rMobile stored carbon respiration rate (Log $_{10}$) -6 -0.5 $-1.5, 0.5$ P38Mob _C TrFraction of mobile transfers respired 0 0.1 $0, 0.1$ P39Maint _r Fraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ Scaling parameters -7 -5 2 $0.96, 1.65$ P40Rsoil ₁ Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.62, 1.53$ P42Rsoil ₃ Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P29	GDD ₂ Spring photosynthetic GDD maximum		500	1 000	660,1 000		
P30LitdLitter respiration rate (Log_{10}) -7 -1 $-6.6, -3.7$ P31Litd, T_d Litter respiration temperature dependence 0.001 0.1 $0.01, 0.1$ P32SOM _{Sd} Slow cycling SOM _C respiration rate (Log_{10}) -6 -1 $-4.55, 3.11$ P33SOM _{Sd} T_d Slow cycling SOM _C temperature dependence 0.01 0.2 $0.01, 0.19$ P34SOM _{Fd} Passive cycling SOM _C respiration rate (Log_{10}) -6 -1 $-6.38, -5.15$ P35Rroot _d Root respiration rate (Log_{10}) -6 -1 $-5.09, -3.77$ P36Rroot _d T_d Root respiration rate temperature dependence 0.01 0.2 $0.07, 0.2$ P37Mob _C rMobile stored carbon respiration rate (Log_{10}) -6 -0.5 $-1.5, 0.5$ P38Mob _C rFraction of mobile transfers respired 0 0.1 $0, 0.1$ P39Maint _r Fraction of GPP respired for maintenance 0.1 0.5 2 $0.96, 1.65$ P40Rsoil ₁ Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P41Rsoil ₂ Soil respiration scaling co-efficient (data set 2) 0.5 2 $0.45, 1.65$ P42Rsoil ₃ Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$				Respiratio	n parameters			
P31Litd T_d Litter respiration temperature dependence0.0010.10.01, 0.1P32SOM _{sd} Slow cycling SOM _C respiration rate (Log ₁₀)-6-1-4.55, 3.11P33SOM _{sd} T _d Slow cycling SOM _C temperature dependence0.010.20.01, 0.19P34SOM _{Pd} Passive cycling SOM _C respiration rate (Log ₁₀)-6-1-6.38, -5.15P35Rroot _d Root respiration rate (Log ₁₀)-6-1-5.09, -3.77P36Rroot _d T _d Root respiration rate temperature dependence0.010.20.07, 0.2P37Mob _C rMobile stored carbon respiration rate (Log ₁₀)-6-0.5-1.5, 0.5P38Mob _C TrFraction of mobile transfers respired00.10, 0.1P39Maint _r Fraction of GPP respired for maintenance0.10.50.1, 0.44P40Rsoil ₁ Soil respiration scaling co-efficient (data set 1)0.520.96, 1.65P41Rsoil ₂ Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P30	Lit _d	Litter respiration rate (Log_{10})	-7	-1	-6.6, -3.7		
P32 SOM _{sd} Slow cycling SOM _c respiration rate (Log_{10}) -6 -1 -4.55, 3.11 P33 SOM _{sd} T _d Slow cycling SOM _c temperature dependence 0.01 0.2 0.01, 0.19 P34 SOM _{Pd} Passive cycling SOM _c respiration rate (Log_{10}) -6 -1 -6.38, -5.15 P35 Rroot _d Root respiration rate (Log_{10}) -6 -1 -5.09, -3.77 P36 Rroot _d T _d Root respiration rate temperature dependence 0.01 0.2 0.07, 0.2 P37 Mob _{Cr} Mobile stored carbon respiration rate (Log_{10}) -6 -0.5 -1.5, 0.5 P38 Mob _C r Fraction of mobile transfers respired 0 0.1 0, 0.1 P39 Maint _r Fraction of GPP respired for maintenance 0.1 0.5 0.1, 0.44 Scaling parameters Scaling parameters Scaling parameters Scaling parameters P40 Rsoil ₁ Soil respiration scaling co-efficient (data set 1) 0.5 2 0.62, 1.53 P41 Rsoil ₂ Soil respiration scaling co-efficient (data set 3) 0.5 2 0.45, 1.65	P31	Lit_dT_d	Litter respiration temperature dependence	0.001	0.1	0.01, 0.1		
P33SOMsdTdSlow cycling SOMc temperature dependence 0.01 0.2 $0.01, 0.19$ P34SOMPdPassive cycling SOMc respiration rate (Log10) -6 -1 $-6.38, -5.15$ P35RrootdRoot respiration rate (Log10) -6 -1 $-5.09, -3.77$ P36RrootdTdRoot respiration rate temperature dependence 0.01 0.2 $0.07, 0.2$ P37MobCrMobile stored carbon respiration rate (Log10) -6 -0.5 $-1.5, 0.5$ P38MobCTrFraction of mobile transfers respired 0 0.1 $0, 0.1$ P39MaintrFraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ P40Rsoil1Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P41Rsoil2Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P32	SOM _{Sd}	Slow cycling SOM_C respiration rate (Log ₁₀)	-6	-1	-4.55, 3.11		
P34SOM _{Pd} Passive cycling SOM _C respiration rate (Log_{10}) -6-1-6.38, -5.15P35Rroot _d Root respiration rate (Log_{10}) -6-1-5.09, -3.77P36Rroot _d T _d Root respiration rate temperature dependence0.010.20.07, 0.2P37Mob _{Cr} Mobile stored carbon respiration rate (Log_{10}) -6-0.5-1.5, 0.5P38Mob _C T _r Fraction of mobile transfers respired00.10, 0.1P39Maint _r Fraction of GPP respired for maintenance0.10.50.1, 0.44Scaling parametersP40Rsoil ₁ Soil respiration scaling co-efficient (data set 1)0.520.96, 1.65P41Rsoil ₂ Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P33	$SOM_{Sd}T_d$	Slow cycling SOM _C temperature dependence	0.01	0.2	0.01, 0.19		
P35Rroot_dRoot respiration rate (Log_{10}) -6 -1 $-5.09, -3.77$ P36Rroot_dT_dRoot respiration rate temperature dependence 0.01 0.2 $0.07, 0.2$ P37Mob_CrMobile stored carbon respiration rate (Log_{10}) -6 -0.5 $-1.5, 0.5$ P38Mob_CTrFraction of mobile transfers respired 0 0.1 $0, 0.1$ P39Maint_rFraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ Scaling parametersP40Rsoil_1Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P41Rsoil_2Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P34	SOM _{Pd}	Passive cycling SOM_C respiration rate (Log ₁₀)	-6	-1	-6.38, -5.15		
P36Rroot_d T_dRoot respiration rate temperature dependence 0.01 0.2 $0.07, 0.2$ P37Mob_CrMobile stored carbon respiration rate (Log ₁₀) -6 -0.5 $-1.5, 0.5$ P38Mob_CTrFraction of mobile transfers respired 0 0.1 $0, 0.1$ P39MaintrFraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ P40Rsoil_1Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P41Rsoil_2Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P35	Rroot _d	Root respiration rate (Log_{10})	-6	-1	-5.09, -3.77		
P37Mob_CrMobile stored carbon respiration rate (Log_{10}) -6 -0.5 $-1.5, 0.5$ P38Mob_CTrFraction of mobile transfers respired0 0.1 $0, 0.1$ P39MaintrFraction of GPP respired for maintenance 0.1 0.5 $0.1, 0.44$ Scaling parametersP40Rsoil_1Soil respiration scaling co-efficient (data set 1) 0.5 2 $0.96, 1.65$ P41Rsoil_2Soil respiration scaling co-efficient (data set 2) 0.5 2 $0.62, 1.53$ P42Rsoil_3Soil respiration scaling co-efficient (data set 3) 0.5 2 $0.45, 1.65$	P36	$\operatorname{Rroot}_{d}T_{d}$	Root respiration rate temperature dependence	0.01	0.2	0.07, 0.2		
P38Mob _C T _r Fraction of mobile transfers respired00.10, 0.1P39Maint _r Fraction of GPP respired for maintenance0.10.50.1, 0.44Scaling parametersP40Rsoil ₁ Soil respiration scaling co-efficient (data set 1)0.520.96, 1.65P41Rsoil ₂ Soil respiration scaling co-efficient (data set 2)0.520.62, 1.53P42Rsoil ₃ Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P37	Mob _{Cr}	Mobile stored carbon respiration rate (Log_{10})	-6	-0.5	-1.5, 0.5		
P39MaintrFraction of GPP respired for maintenance0.10.50.1, 0.44P40Rsoil1Soil respiration scaling co-efficient (data set 1)0.520.96, 1.65P41Rsoil2Soil respiration scaling co-efficient (data set 2)0.520.62, 1.53P42Rsoil3Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P38	Mob _C T _r	Fraction of mobile transfers respired	0	0.1	0, 0.1		
P40Rsoil1Soil respiration scaling co-efficient (data set 1)0.520.96, 1.65P41Rsoil2Soil respiration scaling co-efficient (data set 2)0.520.62, 1.53P42Rsoil3Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P39	Maint.	Fraction of GPP respired for maintenance	0.1	0.5	0.1, 0.44		
P40Rsoil1Soil respiration scaling co-efficient (data set 1)0.520.96, 1.65P41Rsoil2Soil respiration scaling co-efficient (data set 2)0.520.62, 1.53P42Rsoil3Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65		ĩ	1	Scaling pa	rameters			
P41Rsoil2Soil respiration scaling co-efficient (data set 2)0.520.62, 1.53P42Rsoil3Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P40	Rsoil ₁	Soil respiration scaling co-efficient (data set 1)	0.5	2	0.96, 1.65		
P42Rsoil_3Soil respiration scaling co-efficient (data set 3)0.520.45, 1.65	P41	Rsoil ₂	Soil respiration scaling co-efficient (data set 2)	0.5	2	0.62, 1.53		
	P42	Rsoil ₃	Soil respiration scaling co-efficient (data set 3)	0.5	2	0.45, 1.65		

$$J = \left(\sum_{i=1}^{M} j_i\right) / M,\tag{2}$$

where M is the number of data streams used.

Thus, each individual cost function is averaged by the number of observations, and the average of the cost functions from all data streams is taken as the total cost function. In this manner, each data stream is given equal importance in the optimization (Franks *et al.*, 1999; Barrett *et al.*, 2005).

Model benchmarking – ANN ensemble

We used an ANN to benchmark the FöBAAR model performance (e.g., Abramowitz *et al.*, 2007) and characterize the climatic sensitivity of ecosystem-atmosphere carbon exchange. An ANN is an inductive modeling approach based on statistical multivariate modeling (Bishop, 1995; Rojas, 1996) by which one can map drivers directly onto observations (e.g., Moffat *et al.*, 2010). The benchmarking framework used in this paper is based on a feed-forward ANN with a sigmoid activation function trained with a back propagation algorithm (Moffat *et al.*, 2010). An ensemble of six ANNs was trained on nongap-filled eddy-covariance carbon fluxes only. It should be noted that the ANN is a benchmark only for short-term environmental controls on hourly NEE, as it does not account for lagged effects on ecosystem state or function, or long-term changes in pool sizes.

The ANN was also used as a gap-filling tool to compare the gap-filled eddy-covariance carbon fluxes. When used as a gap-filling tool (e.g., Moffat *et al.*, 2007), the ANN was trained on each year of eddy-covariance carbon flux data separately. Thus applied, the ANN agreed with the annual carbon flux from the independently gap-filled data with a root mean square error of 32 g C m⁻².

Experimental set-up

We divided the 18 years of available data into three distinct 6 year periods (1992–1997; 1998–2003; 2004–2009; Fig. 2) to

perform two experiments. In the first experiment, we used the middle period (Period 2, Fig. 1) to quantify the added benefit of using different data streams as constraints. This involved optimizing FöBAAR using as constraints either: (1) only hourly NEE data, (2) hourly, monthly, and yearly NEE data, or (3) all eddy-covariance carbon flux data (hourly, monthly, yearly) and ancillary data (Table 1). We then assessed the optimized model performance for the two periods not used for training. The ANN was trained to the eddy-covariance carbon flux data for the same 6 year period on which the FöBAAR model.

The second experiment was designed to test whether model deficiencies highlighted by the first experiment could be resolved by training the model on each period. In the second experiment, we used all available data to optimize the FöBAAR model on each 6 year period individually. This allowed us to assess changes in model parameters when optimized on different periods.

Finally, for each of the three approaches to constraining the model (1, 2, and 3 above) in the first experiment, we projected carbon stocks and fluxes to 2100, to assess the effect of each constraint approach on the future propagation of uncertainty.

Downscaled future climate projections

For the climate change projection, we used downscaled data (Hayhoe *et al.*, 2007) from the regionalized projection of the GFDL-CM global coupled climate-land model (Delworth *et al.*, 2006) driven with socioeconomic change scenario A1FI (Denham KL *et al.*, 2007). Model projections for Harvard forest under this scenario predict an increase in atmospheric CO₂ to 969 ppm by 2100 and an increase in mean annual temperature from 7.1 to 11.9 °C.

Results

Assessing the benefit of additional constraints

We first tested the benefit of using flux and ancillary data for constraining model projections. Here, we use



Fig. 1 Model uncertainty for NEE, GPP, Ra, and Rh, for the FöBAAR model. The FöBAAR model was constrained on data in Period 2 and tested on Periods 1 and 3. Three different approaches to constraining the model are shown: (1) using all data available (flux and biometric, black), (2) using hourly tower measurements of NEE, and monthly and annual aggregates (dark gray), and (3) using only hourly tower measurements of NEE (light gray). The shaded areas thus represent the confidence in model projections, without a direct comparison to data.

the middle six years of the time series (Period 2, Fig. 2) to optimize the FöBAAR model and the other two periods for testing, assessing three different approaches to constraining the model (see Materials and methods section). When using only hourly NEE as a constraint, uncertainty in annual mean NEE model estimates was large (± 200 g C m⁻² yr⁻¹ 95% CI, Fig. 1). Particularly large uncertainty was evident among the component fluxes of gross primary productivity (± 320 g C m⁻² yr^{-1}), autotrophic (±410 g C m⁻² yr⁻¹) and heterotrophic respiration (± 290 g C m⁻² yr⁻¹). The use of monthly and annual flux aggregates largely reduced uncertainty in model estimates of annual NEE (to ± 60 g $C m^{-2} yr^{-1}$) during both the training and test periods, though only slightly reduced equifinality, shown in Fig. 1 as relatively large uncertainties in the component fluxes. Using all available data to constrain the model only slightly reduced uncertainty for annual flux estimates but gave a large reduction in uncertainty in the responsible processes (Fig. 1). Uncertainty in modeled fluxes in the test periods was comparable to that in the training period for each of the constraint approaches.

FöBAAR and ANN evaluation in training and test periods

In the following analysis, we trained both FöBAAR using all constraints and the ANN on Period 2 using only short-term flux constraints (Fig. 2), and tested the models on the other two periods. When trained on Period 2, neither FöBAAR nor the ANN captured the large increase in annual NEE during Period 3 (Fig. 3). The mean annual NEE estimated from the gap-filled tower data for the last 6 years of the time series (Period 3,



Fig. 2 Measured (line) and modeled (light gray area) annual NEE with the FöBAAR model trained on data from Period 2. Horizontal dark gray bars represent measured means for each period.

Fig. 2) was roughly twice that of the previous 6 year period (Period 2, Fig. 2). In contrast, both FöBAAR and the ANN mean annual NEE for Period 3 were comparable with that of Period 2 (Fig. 2). As with all models that do not consider dynamic vegetation, FöBAAR and ANN predictions of NEE outside the training period make the implicit assumption that the climatic sensitivity of ecosystem function does not change between years. Long-term temporal trends in the residuals between the modeled and observed annual NEE can be interpreted as an alteration in the carbon uptake of the ecosystem that is independent of recent changes in the climate variables included in the model. Long-term trends in Harvard forest mean annual uptake [increased by ~ 300 g C m⁻² ($\sim 150\%$) between Period 1 and Period 3] were thus shown to be independent of any recent changes in climate drivers included here.

In general, when trained on Period 2, the FöBAAR model reproduced the mean values for the ancillary data streams, but not the interannual variability. FöBAAR-modeled carbon in wood for Period 2 was well simulated with an RMSE of 51 g C yr⁻¹ (Table 3). Mean annual wood increments were also well captured, allowing for the accurate reproduction of biomass accumulation. Outside of the training period, RMSE performance for woody biomass was reduced, most noticeably for mean annual woody increment in Period 3, where the model under-predicted growth. Interannual variability in modeled wood increment did not show a significant correlation with the observations



Fig. 3 The daily NEE residuals (modeled-measured, g C $m^{-2} day^{-1}$) for FöBAAR and the ANN, showing the seasonal cycle of data-model mismatch, when both models are trained on Period 2. The residuals are shown in polar plots, where a full circle corresponds to 1 year, and monthly intervals are represented by the initial letter of the month. The zero residual is indicated by the inner black circle (solid line). The smoothed line (red, solid) is a 7 day moving average mean based on all years of data in each period.

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	Period 1 (test)		Period 2 (trained)		Period 3 (test)		Period 3 (trained)	
	r^2	RMSE	r^2	RMSE	r^2	RMSE	r^2	RMSE
ANN								
NEE day	0.77	0.17	0.74	0.19	0.76	0.22		
NEE night	0.11	0.10	0.17	0.10	0.19	0.10		
NEE annual	ns	118.18	ns	73.40	ns	213.80		
FöBAAR								
NEE day	0.79	0.16	0.76	0.19	0.75	0.25	0.78	0.20
NEE night	0.09	0.11	0.15	0.11	0.10	0.11	0.14	0.11
NEE annual	ns	63.23	ns	90.57	ns	298.27	ns	87.3
Soil respiration	ns	ns	0.90	0.68	0.71	1.17	0.70	1.08
Leaf area index	0.89	0.86	0.89	0.49	0.76	0.85	0.84	0.71
Litter fall	ns	ns	ns	11.58	ns	50.56	ns	13.34
Woody biomass	1.00	60.15	0.96	52.93	0.99	111.44	0.99	56.08
Woody increment	ns	0.01	ns	0.06	ns	0.15	ns	0.02
Bud burst	0.20	4.24	0.24	4.17	0.21	3.70	0.32	0.57
Leaf drop	0.17	5.74	0.35	3.42	0.18	3.68	0.18	3.68
FöBAAR vs. ANN								
NEE day	0.76	0.18	0.76	0.18	0.71	0.21		
NEE night	0.62	0.06	0.63	0.05	0.54	0.06		
NEE annual	ns	79.18	ns	70.58	ns	80.70		

Table 3 Performance metrics for all data streams used in the FöBAAR model, and net ecosystem exchange for the ANN. See Table 1 for a description of the data used. All nonzero r^2 values are significant for P < 0.05; ns \geq no significant relation found

in any period (Table 3). For canopy processes, the seasonal evolution of LAI was well captured during the training period (r^2 : 0.89, RMSE: 0.49 m² m⁻²). Mean bud-burst dates were well simulated (RMSE: 4.17 days), though interannual variability was not (r^2 : 0.24). Mean leaf senescence was simulated with a similar accuracy (RMSE: 3.4 days) though model correlation with inter-annual variability in senescence was low (r^2 : 0.35). Outside of the training period, model skill at reproducing observations of LAI and phenology declined (Table 3), most notably in Period 3, and in particular for inter-annual variability in leaf senescence. The magnitude of leaf litterfall was well simulated for the training period (RMSE: 12 g C m⁻²) but much less so for Period 3 (RMSE: 51 g C m⁻²), and interannual variability was poorly captured in all three periods.

For hourly daytime NEE in the training period, FöBAAR and the ANN performed comparably (r^2 : 0.76, 0.74), with an equivalent RMSE (0.19). The ANN showed better data-model agreement for the night-time fluxes than the FöBAAR model (Table 3). Cumulative annual fluxes show that both models tended to slightly underestimate the total annual NEE. Neither the FöBAAR model nor the ANN captured the high uptake seen in 2001 (data not shown), suggesting that the observed uptake in this year was not driven by the climatic variables included in this study. The ANN residuals showed no seasonal bias during the training period, whereas the optimized FöBAAR was slightly biased toward underestimating uptake during the growing period, and underestimating carbon released by the ecosystem during winter months (Fig. 3).

For the testing periods, both the ANN and the FöBAAR model performed well for hourly NEE fluxes during 1992–1997 (Period 1, Fig. 3), with no systematic temporal biases (Fig. 3). During 2004–2009, FöBAAR and the ANN both showed strong systematic biases, but only during the growing season (Period 3, Fig. 3) in particular during the months of June, July, August, and September. The correlation of measured and ANN/FöBAAR-modeled day-time NEE for the 2004–2009 period was equivalent to that of the other two periods, but a larger bias was evident for hourly predictions which accumulated to a large bias in the annual total (Table 3). This shows that good correlation to short-term fluxes does not eliminate the possibility of large bias at longer time scales.

Model extrapolation in time

With a perfect understanding of the system, a model trained on one period should be able to predict the fluxes in the other periods. Experiment 1 showed that neither model used here could do so at Harvard forest. In Experiment 2, we calibrated the FöBAAR model to each period individually. When calibrating FöBAAR to all of the available data on the three individual periods, little bias is evident for FöBAAR NEE during that

period, but large biases are evident in the other periods (Fig. 4). Calibrating to the whole time series thus overestimates annual NEE for the first period, gives low bias in annual NEE for the middle period, and underestimates annual NEE for the last period. Inter-annual variability in NEE was not captured by the model when trained on any period. Long-term changes in estimated modeled canopy photosynthetic potential (here Vc_{max} , P19) were needed to reproduce the observations. Reproducing the required trend in NEE required an increase in Vc_{max} of ~50% over the 18 years (Fig. 5).



Fig. 4 The cumulative daily NEE residuals (modeled-measured, g C m⁻²) for FöBAAR when trained on each period individually and tested on the other two periods. The red line represents the mean cumulative residual for each 6-year period, and the gray area is one standard deviation about the mean. The dashed black line represents the zero residual.



Fig. 5 The covarying posterior distribution of Vc_{max} and the proportion of gross primary productivity (GPP) respired for maintenance, for the FöBAAR model calibrated independently on each of the three 6-year periods (Fig. 2). Contour lines represent the mean annual GPP (g C m⁻² yr⁻¹) for a particular combination of parameters.

 Vc_{max} co-varied strongly with the proportion of assimilate lost through maintenance respiration (Fig. 5). Such parameter equifinality could explain previous findings that models with very different Vc_{max} values can give comparable estimates of canopy photosynthesis (e.g., Keenan *et al.*, 2011b). Although the use of multiple constraints allowed for the constraining of 24 of the 42 free model parameters, no other significant changes in parameters could be detected between the different periods.

Long-term changes at Harvard forest

From a carbon accounting perspective, changes in the measured annual increment in aboveground biomass over the 18 years (Period 1: ~ 100 g C m⁻²; Period 2: 185 g C m⁻²; Period 3: 220 g C m⁻²) do not fully account for the observed increase in ecosystem carbon storage (NEE). In Period 2, measured aboveground biomass increment was 72% of all carbon sequestered. In Period 3, biomass increment accounted for 42% of observed carbon sequestered. In our model system, which accurately reproduced the mean biomass increment for each period, the remaining increase in uptake could only accumulate in the litter, root, or soil pools. In the model, any increase in the root, litter, or microbial pools would cause an observable increase in soil respiration, yet no increase in soil respiration was observed between the different periods. As the only viable alternative, the model predicted that the remaining uptake (after discounting for increases in aboveground biomass) accumulated in the slow cycling carbon pool at a rate of 300 g C m⁻² yr⁻¹ during Period 3. This contrasted with the accumulation rate of ~70 g $C m^{-2} yr^{-1}$ in Periods 1 and 2. This implies that the reported large increase in net ecosystem carbon uptake, if true, should be detectable in the slow cycling carbon pool.

Ecological forecasting

Long-term model projections of future carbon cycling and stocks (using posterior parameter distributions from the FöBAAR model optimized on Period 2) were strongly dependent on the data used to constrain the model (Fig. 6). The use of short-term (hourly) NEE flux data alone, although it gave a good fit to available hourly NEE measurements (Table 3), led to poor constraint of the long-term evolution of the carbon sinksource state of the forest. Future projections of annual NEE were highly uncertain and ranged from ~ 600 to $-900 \text{ g C m}^{-2} \text{ yr}^{-1}$ (90% CI) in the last decade of the century, compared with an average range of -50 to $-520 \text{ g C m}^{-2} \text{ yr}^{-1}$ (90% CI) in present day conditions



Fig. 6 FöBAAR model projections to 2100 for carbon fluxes (top, g C m⁻² yr⁻¹) and pools (bottom, kg C m⁻²) from 2000 to 2100, using posterior parameters from a model optimization using: (1) only hourly net ecosystem exchange fluxes (dark gray); (2) hourly, monthly, and annual net ecosystem exchange fluxes (medium gray); (3) all flux and ancillary data (light gray) (Table 3). Shaded areas represent 90% confidence limits on model projections, generated by parameter sets taken from the posterior parameter distribution.

(when using only hourly NEE flux data). Largest uncertainty propagated beyond 2050. Uncertainty in autotrophic respiration increased by ~50% by the end of the century and uncertainty in heterotrophic respiration doubled.

The use of long-term (monthly and annual) flux constraints greatly reduced future flux uncertainty. For example uncertainty in future NEE was reduced to within a range of -50 to -450 g C m⁻² yr⁻¹. The largest reduction in uncertainty came from the synchronous use of all data constraints available. The additional use of biometric constraints particularly reduced endogenous uncertainty in future projections of all carbon stocks. With the use of all data constraints, uncertainty in projections of all future stocks and fluxes was within present day uncertainty, with the exception of the slow cycling carbon pools (soil organic matter and carbon in wood). Interestingly, projected future carbon sequestration under climate change is never predicted to increase to the extent observed in the last 18 years at Harvard forest.

Discussion

High-frequency eddy-covariance measurements of forest-atmosphere carbon exchange contain a wealth of information, which can be used to characterize an ecosystems response to climatic drivers, and the evolution of that response over time. When used to constrain a terrestrial carbon cycle model, a large improvement in posterior vs. prior model performance can be achieved for high-frequency fluxes (e.g., Medvigy et al., 2009), along with a reduction in the posterior uncertainty of some model parameters (e.g., Braswell et al., 2005). The annual carbon balance of an ecosystem, however, is not an instantaneous response to a driver, but an accumulation of ecosystem responses to climate variability within the year (le Maire et al., 2010). Here, we show that when using only high-frequency measurements of NEE, small high-frequency model biases can accumulate to give large uncertainty in the total modeled annual carbon balance of the ecosystem over annual and inter-annual time periods. The resulting uncertainty range is of a similar magnitude to the range among models reported from model inter-comparison studies (Heimann et al., 1998; Cramer et al., 2001; Schwalm et al., 2010; Keenan et al., in press). By incorporating information on long-term (monthly, annual) cumulative fluxes into the model optimization, we greatly reduced the uncertainty in model estimates of the annual carbon budget of the forest in both training and test periods.

This reduction was not as pronounced, however, for the components of the carbon budget. When using only eddy-covariance carbon flux data, modeled gross primary productivity and ecosystem respiration compensated for each other to give the observed value for NEE. Such equifinality (Beven, 2006) between quantities allows for large uncertainty in both, but good model performance for the net value of ecosystem carbon exchange. The use of additional constraints in conjunction with eddy-covariance carbon flux data led to a reduction in uncertainty in the component parts of NEE during the test and training periods, if not in NEE itself. In particular, the additional use of biometric and soil flux constraints led to a halving of uncertainty in heterotrophic respiration, and a large reduction in uncertainty regarding the size of the carbon pools.

Synchronously using 15 different data streams as constraints successfully reduced posterior uncertainty in 24 of 42 parameters. The well-constrained nature of the model was evidenced by the accurate simulation of multiple compartments of the ecosystem at various different time scales. Previous model-data fusion efforts have focused on using one or two constraints (with some notable exceptions, e.g., Xu et al., 2006; Medvigy et al., 2009; Richardson et al., 2010; Ricciuto et al., 2011; Weng & Luo, 2011), which invariably led to a low number of constrainable parameters (e.g., ~4 to >6 parameters, Wang et al., 2007; Knorr & Kattge, 2005). Here, constrained parameters were typically associated with processes for which data was available. For instance, the soil organic matter and wood carbon initial pools were well constrained by the measurement data, while the canopy carbon reserve pool was not constrained, as no measurements of mobile canopy carbon were included. Five additional parameters, which were not well constrained, demonstrated strong co-variance with other parameters, thus giving information as to their true distribution. Vcmax and the proportion of recent assimilate used for maintenance respiration serve as a good example in this study – where higher Vc_{max} was compensated for by higher maintenance respiration (Fig. 5). It should be noted that the absolute values of Vc_{max} reported here are specific to the model used. Different assumptions regarding the distribution of light and temperature within the canopy affect the value of Vc_{max} needed to reproduce the observed fluxes (e.g., Keenan et al., 2011b), potentially along with the value assumed for the proportion of assimilate lost to maintenance respiration as shown here. The increased use of multiple data streams in the future will help better constrain models and aid our understanding of long-term processes. However, not all additional data constraints give the same reduction in model uncertainty (Richardson et al., 2010; Ricciuto *et al.*, 2011). In this study, components of ecosystem carbon cycling most uncertain after the integration of all available data were related to gross primary productivity, and the timing and magnitude of above-ground growth and maintenance respiration. Identifying which additional data would better inform model projections should be a focus of future efforts.

By testing the optimized process-based model against the ANN, we have shown that process-based models can reproduce observed NEE measurements as well as data-mining tools. This shows that parsimonious model structures are sufficient to reproduce the observed short-term variability represented in eddycovariance carbon flux data. It also suggests that although eddy-covariance fluxes undoubtedly contain more information than any other individual data constraint, they are not sufficient to adequately test many aspects of more complex models (e.g., Medvigy et al., 2009; Zaehle & Friend, 2010; Bonan et al., 2011). As in other studies (e.g., Hanson et al., 2004; Braswell et al., 2005; Siqueira et al., 2006; Richardson et al., 2007; Urbanski et al., 2007; Richardson et al., 2010; Keenan et al., in press; but see Desai, 2010), the process-based model failed to accurately reproduce observed inter-annual variability in carbon cycling and biomass increments, even within the training period. As the process-based model here was optimized to the data, parameter error can be discounted, leaving model structural error, biotic effects, or missing drivers (e.g., diffuse radiation: Moffat *et al.*, 2010) as potential culprits for the poor model performance for inter-annual variability. Lagged effects of climate variability on ecosystem state (e.g., Gough et al., 2009) have been shown to affect model performance on interannual timescales (Keenan et al., in press), potentially due to inaccurate model allocation structures (Gough et al., 2009). Though it has been suggested that process-based models may effectively reproduce inter-annual variability (Desai, 2010; but see Keenan et al., in press), both biotic and abiotic factors are known to affect normal between-year variability (Richardson et al., 2007). Further work on model structural error, biotic effects, and the impact of unaccounted for drivers should improve our ability to accurately model interannual variability in terrestrial carbon cycling in the future.

Eddy-covariance measurements at Harvard forest suggest a long-term trend of increasing uptake over the 1992–2009 period, with a particularly pronounced increase in uptake in the last 6 years. Results here suggest that long-term changes evidenced by the eddycovariance carbon flux data are independent of recent changes in climate variables included in this study. By comparing the temporal distribution of model-data residuals, we found that nonclimate driven change in carbon fluxes is only evident during the growing season. By comparing the posterior parameters for the FöBAAR model optimized on three separate 6 year periods of contrasting uptake, we show that even with increased leaf area, substantial increases in canopy productivity (here Vc_{max}) are needed to reproduce the observed fluxes.

Although carbon in wood, leaf area and litter-fall all exhibit increases over the past 18 years, a large proportion of the estimated increased uptake is unaccounted for in the measured carbon stocks. Our model results suggest that the rate of accumulation of slow cycling soil organic matter doubled in Period 3 compared with the two earlier periods. Under that working hypothesis, the large influx of carbon in recent years should therefore be detectable with an appropriate sampling intensity (Fernandez et al., 1993) in soil organic matter measurements, with largest increases in the slow cycling soil carbon pool. Without adequate measurements, our model results regarding the fate of the sequestered carbon should not be regarded as strong evidence, and provide but a testable hypothesis. Current efforts to quantify age and residence times of soil carbon with techniques such as isotopic analysis and radiocarbon dating should aid in identifying the ultimate fate of the sequestered carbon.

Inventory data reports an increase in the biomass of Red Oak within the tower footprint (~20% increase over the last 18 years), and a concurrent increase in Red Oak leaf area. Other species in the footprint of the tower do not show a comparable increase, with the exception of a slight increase in understory Hemlock. Changes in community dynamics provide one potential explanation of the changes in ecosystem uptake. Increasing understory activity has been suggested to have the potential to explain trends (Jolly et al., 2004), through enhanced photosynthetic uptake before the overstory canopy has developed in spring, or after it has senesced in autumn. Understory activity, however, is unlikely to explain the consistent higher uptake throughout the season as observed here. The observed increase in forest carbon uptake could also be due to higher atmospheric CO₂ levels (Cramer et al., 2001), or the cumulative effect of nitrogen deposition. Farquhar et al. (1980) photosynthesis model used in this study accounts for effects of increased atmospheric carbon, though there is significant uncertainty as to the direct effect of carbon fertilization (e.g., Long et al., 2006). Although nitrogen deposition at Harvard forest is 10-20 times above historic background levels (http:// www.chronicn.unh.edu/), it remains only ~12% of annual N mineralization (Munger et al., 1998), and control data from long-term nitrogen fertilization studies do not report a significant increase in foliar nitrogen

(data not shown). It should be noted that there is no evidence to suggest that any of the processes discussed above could, in isolation, realistically lead to a \sim 50% increase in the photosynthetic potential of the canopy.

Future projections from terrestrial models have been reported to diverge greatly under climate change (Friedlingstein et al., 2006; Heimann & Reichstein, 2008). Such divergence could be explained by process misparameterization, or misspecification. We show that using short-term high-frequency eddy-covariance carbon flux data alone to inform model parameterization allows for divergent future projections, even with good model performance when tested against current data. Parameter misspecification could therefore potentially explain the different future trajectories reported by different models. We show that using orthogonal constraints can reduce this divergence, leading to a better datainformed model projection. Using long-term flux data in combination with biometric data greatly reduced endogenous (internal to the model system) uncertainty in predictions of how net carbon sequestration at Harvard forest would respond to future climate change. Considerable uncertainty in the components of NEE remained, due to equifinality between gross photosynthesis and autotrophic respiration.

Although process-based models should theoretically be more reliable than empirical models under future climate scenarios (see Keenan *et al.*, 2011a for discussion), not all processes are fully understood (e.g., species adaptation, down-regulation, nitrogen cycling). Such exogenous uncertainty is shown here to be large, with the optimized model incapable of reproducing the observed long-term trend in carbon cycling at Harvard forest without temporal changes in parameters. This suggests that, when the model is sufficiently informed by data, model process representation still represents a large source of uncertainty for making future projections, making the statistical uncertainty in ecological forecasts an underestimate of the true uncertainty.

Models of forest carbon cycling, such as the one used here, have been coupled with earth-system models to project terrestrial carbon sinks and sources (e.g., Sitch *et al.*, 2008) and feedbacks to climate change in the 21st century (Cox *et al.*, 2000; Fung *et al.*, 2005; Friedlingstein *et al.*, 2006). Results have been incorporated into the assessment reports of the Intergovernmental Panel on Climate Change (Denham KL *et al.*, 2007) to guide mitigation efforts by governments and public (Solomon *et al.*, 2007), though models diverge largely when projecting the future responses to climate change (Friedlingstein *et al.*, 2006; Denham KL *et al.*, 2007). None of the terrestrial carbon cycle models used, however, are directly informed by data. Here, we have shown how this can lead to overconfidence in individual model projections. Model intercomparison studies that use data-informed models would be a significant step toward rigorously assessing errors due to model process representation, and improving our ability to provide policy-actionable predictions of future carbon cycle responses to change.

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